

J. O. Metzger, *Angew. Chem.* **2000**, *112*, 2898; *Angew. Chem. Int. Ed.* **2000**, *39*, 2758.

- [14] For the optical rotation of **8**, see J. A. Bajgrowicz, A. El. Hallaoui, R. Jacquier, C. Pigiere, P. Viallefont, *Tetrahedron* **1985**, *41*, 1833; for details on the determination of the stereochemistry of several of the products see the Supporting Information.
- [15] The potential for the formation of a five-membered chelate also exists (coordination of the metal with the amide nitrogen atom and the ester carbonyl group). The observed sense of stereoinduction is not consistent with formation of a tetrahedral five-membered chelate. The reactions with **14** provide additional support for a seven-membered chelate.
- [16] A. Kubo, H. Kubota, M. Takahashi, K. Nunami, *J. Org. Chem.* **1997**, *62*, 5830.

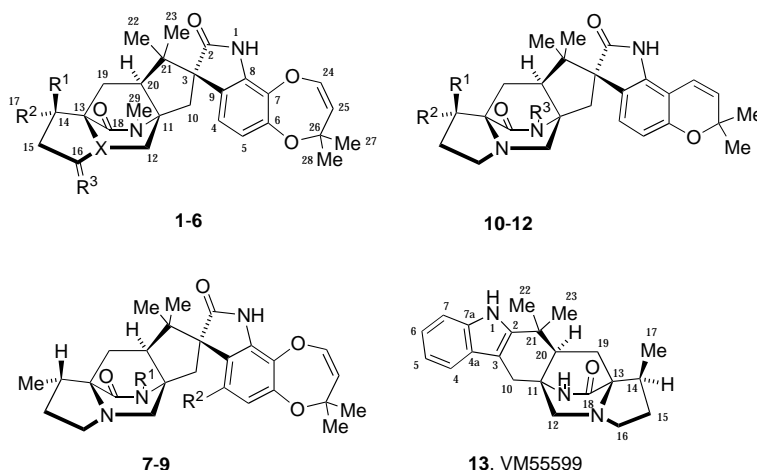


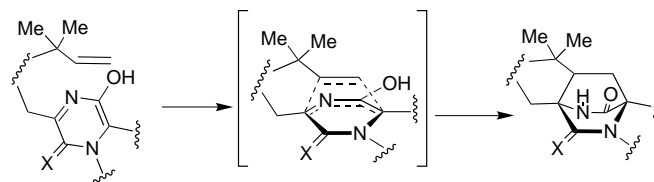
Figure 1. **1**, paraherquamide A: $R^1 = \text{OH}$, $R^2 = \text{Me}$, $R^3 = \text{H}_2$, $X = \text{N}$; **2**, paraherquamide B: $R^1 = \text{H}$, $R^2 = \text{H}$, $R^3 = \text{H}_2$, $X = \text{N}$; **3**, paraherquamide C: $R^1 = R^2 = \text{CH}_2$, $R^3 = \text{H}_2$, $X = \text{N}$; **4**, paraherquamide D: $R^1 = \text{O}$, $R^2 = \text{CH}_2$, $R^3 = \text{H}_2$, $X = \text{N}$; **5**, VM55596: $R^1 = \text{OH}$, $R^2 = \text{Me}$, $R^3 = \text{H}_2$, $X = \text{N}^+ - \text{O}^-$; **6**, VM55597: $R^1 = \text{OH}$, $R^2 = \text{Me}$, $R^3 = \text{O}$, $X = \text{N}$; **7**, paraherquamide E (VM54159): $R^1 = \text{Me}$, $R^2 = \text{H}$; **8**, SB203105: $R^1 = \text{Me}$, $R^2 = \text{OH}$; **9**, SB200437: $R^1 = \text{H}$, $R^2 = \text{H}$; **10**, paraherquamide F (VM55594): $R^1 = \text{H}$, $R^2 = \text{Me}$, $R^3 = \text{Me}$; **11**, paraherquamide G (VM54158): $R^1 = \text{OH}$, $R^2 = \text{Me}$, $R^3 = \text{Me}$; **12**, VM55595: $R^1 = \text{H}$, $R^2 = \text{Me}$, $R^3 = \text{H}$.

Studies on the Biosynthesis of Paraherquamide: Synthesis and Incorporation of a Hexacyclic Indole Derivative as an Advanced Metabolite**

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The paraherquamides (Figure 1),^[1] along with the brevianamides,^[2] marcfortines,^[3] and sclerotamides^[4] are indolic fungal metabolites that share the common structural feature of an unusual bicyclo[2.2.2]diazaoctane core. It has been postulated that the bicyclo[2.2.2]diazaoctane ring system arises through an intramolecular hetero Diels–Alder cycloaddition of the isoprene moiety across the α -carbons of the amino acid subunits, as shown in Scheme 1.^[5]

In 1993, Everett and co-workers isolated a very minor metabolite that also possesses the bicyclo[2.2.2]diazaoctane core, VM55599 (**13**, Figure 1), from *Penicillium* sp. (IMI 332995) which produces paraherquamide A.^[6] Based on



Scheme 1. Proposed formation of the bicyclo[2.2.2]diazaoctane ring system in the paraherquamides.

the structural similarities of these co-metabolites, Everett et al. speculated that VM55599 might be a biosynthetic precursor to paraherquamide A.^[6] The relative stereochemistry of VM55599 as shown in Figure 1 was assigned by ¹H NMR spectroscopy with nuclear Overhauser enhancements but the absolute configuration of this substance remains unknown. The stereochemistry of the methyl group in the β -methylproline ring was assigned as being *syn* to the bridging isoprene moiety. In all other known members of the paraherquamide family, the methyl group in the β -methylproline ring is disposed *anti* to the bridging isoprene moiety. If VM55599 was indeed a precursor to paraherquamide A, then oxidation of the β -methylproline ring would have to occur with inversion of stereochemistry at the C-14 center that bears the methyl group.

Previous studies from this laboratory on the biosynthesis of paraherquamide A demonstrated that L-isoleucine is the precursor to the β -methyl- β -hydroxy proline ring of paraherquamide A.^[7] The relative disposition of the methyl group in the prolyl ring is retained in the biosynthetic conversion of L-isoleucine into paraherquamide A and, thus, the hydroxylation at C-14 occurs with net retention. These findings bring into question the potential intermediacy of VM55599 in the biosynthesis of the paraherquamides. Furthermore, if L-isoleucine is also the precursor to VM55599, then the absolute stereochemistry of this metabolite must be that depicted in

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to increase the miscibility of these substances in the culture broth without inhibiting production of paraherquamide A. Feeding experiments were performed on *P. fellutanum* (ATCC20841) with all four potential precursors, followed by isolation and purification of paraherquamide A. Within the limits of detection by ^{13}C NMR spectroscopy and mass spectrometry no incorporation was observed for VM55599 ((\pm)-**13**) or its oxidized counterpart ((\pm)-**14**). In addition, no incorporation was observed for the diketopiperazine ((\pm)-**16**). However, for the C-14 epimer of VM55599 ((\pm)-**15**), significant incorporation was observed by ^{13}C NMR spectroscopy at C-12 and C-18 of paraherquamide A. From analysis of the electrospray mass spectrum, incorporation was determined to be 0.72 % for the intact doubly labeled material.^[11] ^{13}C -Monolabeled paraherquamide A, from catabolism of ((\pm)-**15**, was not detected in the mass spectrum. The implications of these observations are considerable.

Since the diketopiperazine ((\pm)-**16**) was not incorporated, this raises interesting questions concerning the timing of the reduction of the prolyl-derived carbonyl group. The incorporation of compound ((\pm)-**15** in significant isotopic yield, indicates that the formation of the bicyclo[2.2.2]diazaoctane occurs at the stage with the nonoxidized tryptophyl moiety (that is, the indolyl group). This mandates that oxidations of the indole ring to form both the catechol-derived dioxepin and spirooxindole occur *after* the formation of this intermediate. It thus follows that the dioxepin-derived isoprenylation and the *S*-adenosylmethionine-mediated *N*-methylation reactions occur late in the pathway. These results also cast considerable doubt on the intermediacy of VM55599^[6] and its oxidized precursor **14** in the paraherquamide biosynthesis and provide additional circumstantial evidence that VM55599 is a minor shunt metabolite. Finally, the present work documents the intermediacy of an advanced metabolite **15**, which contains the core structural elements of the paraherquamide framework, prior to a series of oxygenation reactions. Efforts to elucidate the exact sequence of biosynthetic reactions immediately preceding and following the formation of **15** are currently under way in these laboratories.

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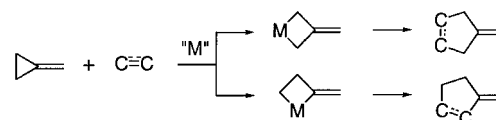
- [1] a) M. Yamazaki, E. Okuyama, M. Kobayashi, H. Inoue, *Tetrahedron Lett.* **1981**, 22, 135–136; b) J. G. Ondeyka, R. T. Goegelman, J. M. Schaeffer, L. Kelemen, L. Zitano, *J. Antibiot.* **1990**, 43, 1375–1379; c) J. M. Liesch, C. F. Wichmann, *J. Antibiot.* **1990**, 43, 1380–1386; d) S. E. Blanchflower, R. M. Banks, J. R. Everett, B. R. Manger, C. Reading, *J. Antibiot.* **1991**, 44, 492–497.
- [2] a) A. J. Birch, J. J. Wright, *J. Chem. Soc. Chem. Commun.* **1969**, 644–645; b) A. J. Birch, J. J. Wright, *Tetrahedron* **1970**, 26, 2329–2344; c) A. J. Birch, R. A. Russell, *Tetrahedron* **1972**, 28, 2999–3008; d) B. A. Bird, A. T. Remaley, I. M. Campbell, *Appl. Environ. Microbiol.* **1981**, 42, 521–525; e) B. A. Bird, I. M. Campbell, *Appl. Environ. Microbiol.* **1982**, 43, 345; f) J. E. Robbers, J. W. Straus, *Lloydia* **1975**, 38, 355; g) R. R. M. Paterson, D. L. Hawksworth, *Trans. Br. Mycol. Soc.* **1985**, 85, 95–100; h) B. J. Wilson, D. T. C. Yang, T. M. Harris, *Appl. Microbiol.* **1973**, 26, 633–635; i) J. Coetzer, *Acta Crystallogr. Sect. B* **1974**, 30, 2254–2256.
- [3] a) J. Polonsky, M.-A. Merrien, T. Prange, C. Pascard, *J. Chem. Soc. Chem. Commun.* **1980**, 601–602; b) T. Prange, M.-A. Buillion, M.

- Vuilhorgne, C. Pascard, J. Polonsky, *Tetrahedron Lett.* **1980**, 22, 1977–1980.
- [4] A. C. Whyte, J. B. Gloer, *J. Nat. Prod.* **1996**, 59, 1093–1095.
- [5] a) A. E. A. Porter, P. G. Sammes, *J. Chem. Soc. Chem. Commun.* **1970**, 1103; b) R. M. Williams, E. Kwast, H. Coffman, T. Glinka, *J. Am. Chem. Soc.* **1989**, 111, 3064–3065; c) R. M. Williams, T. Glinka, E. Kwast, H. Coffman, J. K. Stille, *J. Am. Chem. Soc.* **1990**, 112, 808–821; d) J. F. Sanz-Cervera, T. Glinka, R. M. Williams, *J. Am. Chem. Soc.* **1993**, 115, 347–348; e) J. F. Sanz-Cervera, T. Glinka, R. M. Williams, *Tetrahedron* **1993**, 49, 8471–8472; f) L. R. Domingo, J. F. Sanz-Cervera, R. M. Williams, M. T. Picher, J. A. Marco, *J. Org. Chem.* **1997**, 62, 1662–1667; g) E. M. Stocking, J. F. Sanz-Cervera, R. M. Williams, *Angew. Chem.* **1999**, 111, 880–883; *Angew. Chem. Int. Ed.* **1999**, 38, 786–789; h) E. M. Stocking, R. M. Williams, J. F. Sanz-Cervera, *J. Am. Chem. Soc.* **2000**, 122, 9080–9098.
- [6] S. E. Blanchflower, R. M. Banks, J. R. Everett, C. Reading, *J. Antibiot.* **1993**, 46, 1355–1363.
- [7] a) E. M. Stocking, J. F. Sanz-Cervera, R. M. Williams, C. J. Unkefer, *J. Am. Chem. Soc.* **1996**, 118, 7008–7009; b) E. M. Stocking, R. A. Martinez, L. A. Silks, J. F. Sanz-Cervera, R. M. Williams, *J. Am. Chem. Soc.*, submitted.
- [8] Elucidation of the absolute stereochemistry of VM55599 is in progress. The absolute stereostructures for compounds **13**–**16** (which are racemic) are depicted with the absolute configurations presumed to be those of the natural metabolites.
- [9] a) R. M. Williams, J. F. Sanz-Cervera, F. Sancenon, J. A. Marco, K. Halligan, *J. Am. Chem. Soc.* **1997**, 119, 1090–1091; b) R. M. Williams, J. F. Sanz-Cervera, F. Sancenon, J. A. Marco, K. Halligan, *Bioorg. Med. Chem.* **1998**, 6, 1233–1241; c) E. M. Stocking, J. F. Sanz-Cervera, R. M. Williams, *J. Am. Chem. Soc.* **2000**, 122, 1675–1683.
- [10] See Supporting Information for experimental details and spectroscopic data.
- [11] Calculated according to the method outlined in: J. B. Lambert, H. B. Shurvell, D. A. Lightner, R. G. Cooks, *Organic Structural Spectroscopy*, Prentice Hall, New Jersey, **1998**, pp. 447–448.

Novel [3+2] Cycloaddition of Alkylidenecyclopropanes with Aldehydes Catalyzed by Palladium

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A metal-catalyzed cycloaddition between methylenecyclopropane and a carbon–carbon multiple bond can proceed through two different reaction pathways to give regioisomeric [3+2] carbocycles (Scheme 1).^[1–3] The research groups of



Scheme 1. Metal-catalyzed cycloaddition of methylenecyclopropane and a carbon–carbon multiple bond.

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